NSF-CBMS Regional Conference Series in Probability and Statistics Volume 6

STATISTICAL INFERENCE FROM GENETIC DATA ON PEDIGREES

Elizabeth A. Thompson

University of Washington

Institute of Mathematical Statistics Beachwood, Ohio

American Statistical Association Alexandria, Virginia Conference Board of the Mathematical Sciences

Regional Conference Series in Probability and Statistics

Supported by the National Science Foundation

The production of the *NSF-CBMS Regional Conference Series in Probability and Statistics* is managed by the Institute of Mathematical Statistics: Barry Arnold, IMS Managing Editor, Statistics; Patrick Kelly, IMS Production Editor; Julia Norton, IMS Treasurer; and Elyse Gustafson, IMS Executive Director.

Library of Congress Control Number: 00-134575

International Standard Book Number 0-940600-49-8

Copyright © 2000 Institute of Mathematical Statistics

All rights reserved

Printed in the United States of America

Contents

\mathbf{P}_{1}	refac	e	xi
Ta	able	of Notation	xiii
1	Gen 1.1 1.2 1.3 1.4	nes, Pedigrees and Genetic Models DNA, alleles, loci, genotypes, and phenotypes Mendel's laws and meiosis indicators Pedigrees: the conditional independence structure Models, parameters, and inferences	1 1 3 4 7
2		elihood, Estimation and Testing Likelihood and log-likelihood. Estimation, information, and testing Population allele frequencies The EM algorithm; general formulation Gene counting and the ABO blood types EM estimation for quantitative trait data	 11 11 13 16 20 22 25
3	Ger 3.1 3.2 3.3 3.4 3.5 3.6 3.7 3.8	ne Identity by Descent Kinship and inbreeding coefficients Methods of computation Data on inbred individuals Multi-gamete kinship and gene <i>ibd</i> Patterns of gene <i>ibd</i> in pairs of individuals Observations on related individuals Monte Carlo estimation of expectations Reduction of Monte Carlo variance	 29 30 32 34 36 39 44 46
4	Gen 4.1 4.2 4.3 4.4 4.5	netic Linkage Linkage and recombination: genetic distance Haplotypes, linkage, and association Lod scores for two-locus linkage analysis Power, information and <i>Elods</i> Two-locus kinship and gene identity	53

	4.6	Homozygosity mapping with a single marker
	4.7	Meiosis at multiple linked loci
	4.8	Multi-locus kinship and gene identity
5	Mo	dels for Meiosis 69
	5.1	The meiosis process
	5.2	From chromatids to crossovers
	5.3	From chiasmata to recombination patterns
	5.4	The chiasmata avoidance process
	5.5	Chromatid interference
	5.6	Count-location models for chiasmata
	5.7	Renewal process models of chiasma formation
6	Lil	celihoods on Pedigrees 81
	6.1	The Baum algorithm and "Peeling"
	6.2	Exact likelihoods for multiple markers
	6.3	Computations on large but simple pedigrees
	6.4	Example of peeling a zero-loop pedigree
	6.5	Computations on complex pedigrees
	6.6	Models with Gaussian random effects
7		nte Carlo Estimates on Pedigrees 93
	7.1	Baum algorithm for conditional probabilities
	7.2	An EM algorithm for map estimation
	7.3	Importance sampling for likelihoods
	7.4	Risk probabilities and reverse peeling
	7.5	Elods and SIMLINK
	7.6	Sequential imputation
8	Ma	rkov chain Monte Carlo on Pedigrees 103
	8.1	Simulation conditional on data: MCMC
	8.2	Single-site updating methods
	8.3	Combining exact computation and Monte Carlo
	8.4	Tightly-linked loci: the M-sampler
9	Lik	elihood Ratios for Genetic Analysis 115
	9.1	Monte Carlo likelihood ratio estimation
	9.2	Monte Carlo relative likelihood surfaces
	9.3	Monte Carlo EM for the mixed model
	9.4	Likelihood estimators for complex models
	9.5	Likelihood estimation of gene locations
	9.6	Marker <i>ibd</i> and complete-data log-likelihoods

ii

CONTENTS

10	Cas	e studies using the M- and LM-samplers 129	9
	10.1	Background to a study	9
	10.2	Conditional gene <i>ibd</i> probabilities	1
	10.3	Likelihoods and log-likelihoods	3
	10.4	Gene <i>ibd</i> in a smaller example	5
	10.5	MCMC lod score estimation	7
	10.6	Better MCMC lod scores	0
11	Oth	er Monte Carlo Likelihoods in Genetics 147	7
	11.1	Improving pedigree samplers	7
		Interference by Metropolis-Hastings	
	11.3	Inference of typing or pedigree error	4
	11.4	Other Monte-Carlo procedures for linkage analysis	6
	11.5	Monte-Carlo likelihoods in population genetics	6

iii

CONTENTS

List of Tables

2.1	Conditional and joint probabilities of feasible mother-child genotype combinations	17
2.2	Data and estimated frequencies for Bernstein's analysis of <i>ABO</i> blood type determination	18
2.3	Sequence of EM iterates for the example of estimation of the frequency of a recessive allele	23
2.4	EM iterates for the estimation of ABO allele frequencies. The iterates of allele frequencies, and the resulting conditional probabilities of genotype AO and BO , given phenotypes A and B , respectively, are shown in the upper left panel. Then are shown the resulting expected genotype frequencies, given the observed phenotype frequencies and current allele frequency estimates (E-step). Finally, in the lower right	
	are shown the new iterates of the allele frequencies (M-step)	24
3.1	States of gene <i>ibd</i> among the four genes of two individuals	37
3.2	Values of κ , and kinship coefficient ψ , for some standard relationships between two non-inbred individuals	37
3.3	Gene <i>ibd</i> state probabilities at a single locus for a pair of sisters with an aunt, niece, or half-sib. The states are given in the reduced genotypic state-class form, in which the paternal and maternal genes	40
	of the three individuals are not distinguished	43
4.1	Critical values for a test size $\alpha = 0.025$ and base-10 lod scores for binomial samples	56
4.2	The groups of offspring genotypes in an intercross design. Note the A_1A_1, B_1B_2 type includes both double-heterozygote two-locus genotypes A_1B_1/A_2B_2 and A_1B_2/A_2B_1 . The third group includes the four types heterozygous at one of the two loci: A_1A_1, B_1B_2 ,	
	$A_1A_2, B_1B_1, A_2A_2, B_1B_2 \text{ and } A_1A_2, B_2B_2 \dots \dots \dots \dots$	56
4.3	Probabilities of data observations in an intercross design. Given are the total probabilities of each group of types shown in Table 4.2,	
	under the three alternative hypotheses	57
4.4	Comparison of the information in linkage designs per offspring individual sampled: Kullback Leibler information for testing $\rho = 1/2$	
	as a function of the true value of ρ	58

4.5 4.6	Prior autozygosity probabilities over three linked loci for the final	61
	individual of the pedigree of Figure 3.1	66
	True gene identity by descent simulated on the modified Icelandic pedigree	31
10.2	Conditional probabilities of gene identity by descent given the marker data simulated on the modified Icelandic pedigree. Shown are	
10. 3	probabilities $\times 1000$. For details of the cases $(1)-(4)$, see text 13 Conditional probabilities $(\times 1000)$ of gene <i>ibd</i> among the four <i>C</i> alleles on the pedigree of Figure 10.3, with five equally spaced marker	Z
10.4	loci, M1 to M5, and for a recessive trait unlinked to the markers \dots 13 Conditional probabilities (×1000) of gene <i>ibd</i> among the four C	6
	alleles on the pedigree of Figure 10.3, with five equally spaced marker loci, M1 to M5. The trait is now in the map, midway between M2 and M3	6
10.5	Summary of LM-sampler runs on the example of section 10.5. The penultimate run, designated (*), is the run also used for the results of Figures 10.9 and 10.10. The first column shows the M-sampler run discussed in section 10.5. The runs were done on a DEC alpha workstation 400-233, with 192 MB memory	
11.1	Single-site and joint updating schemes on a pedigree	8
11.2	Probabilities of recombination (r) and non-recombination (n) in four equal marker intervals, under interference models I and II and under	
11.3	the Haldane model of no interference (model 0) $\ldots \ldots \ldots \ldots \ldots \ldots 15$ Gene <i>ibd</i> probabilities (×1000) for single loci, and under no	1
	interference (Haldane model)	2
11.4	Gene <i>ibd</i> probabilities $(\times 1000)$ under the recombination pattern probabilities given for interference models (I) and (II) in Table 11.2. Each run consisted of 10,000,000 whole-meiosis Gibbs/Metropolis updates, and took about 1 hour CPU on a DEC Alpha 400-233	
	work-station with 256MB memory	3

vi

List of Figures

1.1	An example pedigree from Goddard et al. (1996)	5
1.2	Meiosis indicators $S_{\bullet,j}$ determine descent of founder genes, at any given locus j . The indicators $S_{i,j}$ are shown under the offspring individual, while the resulting labeled founder genes are shown within each individual	6
1.3	The conditional independence neighborhood structure on a pedigree: (a) the individual neighborhood, and (b) the haplotype neighborhood. The reference individual (a) or haplotype (b) is dark shaded. The individuals [haplotypes] defining the local dependence structure for the reference individual [haplotype] are light shaded	7
3.1	An example pedigree. The structure is the same as that of Figure 1.1 of section 1.3. The four individuals shaded grey are bilateral ancestors of the final individual	30
3.2	The relationship triangle for non-inbred relatives	38
3.3	The relationship of quadruple-half-first-cousins	39
3.4	Meiosis indicators $S_{\bullet,j}$ determine descent of founder genes, and patterns of gene identity by descent, at any given locus j : see Figure 1.2	40
3.5	Determination of probabilities $\Pr(Y_{\bullet,j} \mid S_{\bullet,j})$. The gene descent pattern is assumed to be that of Figure 1.2, and the pairs of genes are shown, rather than the individuals. Five individuals, shown as dashed circles, are assumed to be observed, with marker genotypes as indicated: see text for details. (a) Only genes present in observed individuals are constrained in type. (b) Two genes in a single	
	observed individual are jointly constrained	42
4.1	Example of recombination in a three-generation family	53
4.2	Examples of (a) phase-known and (b) phase-unknown backcross linkage designs	54
4.3	Multi-locus genetic marker data are available on a pair of sibs, and on a third related individual, who may be an aunt, niece, or half-sister of the pair	60
	of the pair	60

5.1	The processes of mitosis and meiosis, shown for a single pair of homologous chromosomes in the nucleus of a cell of a diploid organism. See text for details	70
5.2	The formation of chiasmata, and the crossovers resulting in the chromosomes of the four offspring gametes. The crossovers occurring are the same as in Figure $5.1(e)$	71
6.1	The conditional independence structure of data, in the absence of genetic interference	82
6.2	Pedigree without loops. Shaded individuals are those for whom phenotypic data are assumed to be available	86
8.1	The conditional independence structure for MCMC sampling	110
9.1	Model parameters for estimation of a location likelihood curve	123
10.1	The modified Icelandic pedigree. The four individuals marked "Aff" are affected. Those shaded black have marker data available at the majority of the 17 marker loci. The affected half-shaded individual is typed at only two of the marker loci	1 3 0
10.2	Expected complete-data log-likelihood components for the simulated data on the modified Icelandic pedigree. Shown are $E_{\gamma_0}(\log_e \Pr(\mathbf{Y} \mid \mathbf{S}) \mid \mathbf{Y})$ (upper curve), and $E_{\gamma_0}(\log_e P_{\gamma}(\mathbf{S}) \mid \mathbf{Y})$ for $\gamma = \gamma_0$ (•, lower curve), and for γ to the left (Δ) and right (+) of γ_0 . The location U denotes unlinked. For additional details see text	134
10.3	Hypothetical phenotypic data assumed at each marker locus on the pedigree of Figure 1.1. The four potentially distinct C alleles are	135
10.4	Marker $(M1 \text{ to } M5)$ and trait (Tr) locations for the example of Figure 10.3. The trait locus is at the midpoint of the $(M2, M3)$ interval, so $d_0 = 12.77cM$ and $\rho_0 = 0.1187$	136
10.5	Exact base-10 location lod scores computed using GENEHUNTER 2. The solid lines correspond to having marker data on five pedigree members, and the broken lines to having marker data on only the final affected inbred individual. In each pair, the upper curve corresponds to a trait allele frequency $q = 0.001$, and the lower to q = 0.05	
10.6	Expected complete-data log-likelihoods with the hypothetical data of Figure 10.3 assumed at each of five equally spaced linked marker	139
10.7	Estimated Monte Carlo location base-10 lod score curve for the hypothetical data of Figure 10.3	39
10.8	Base-10 location score curves for the example of section 10.5 re- estimated, shown also with the exact value	41

viii

LIST OF FIGURES

10.9 Expected complete-data log-likelihoods for the example of	
section 10.5, shown for the penultimate run of Table 10.5. The	
notation is as in Figure 10.2. As in that figure, the contribution	
from penetrance terms is shown separately from that for segregation	
terms	43
10.10Estimated conditional probabilities of recombination in the five map	
intervals for the example of section 10.5, shown for the penultimate	
run of Table 10.5. For details, see text $\ldots \ldots \ldots \ldots \ldots \ldots 1$.44
11.1 A multiplex meiosis consisting of an ancestral chain of four meioses.	
These meioses may be jointly updated. For additional details, see	
	10
$ ext{text}$.40

LIST OF FIGURES

Preface

This monograph is based primarily on material presented at the CBMS Summer Course on **Inferences from genetic data on pedigrees** given at Michigan Technical University, Hougton, Michigan, in July 1999. This monograph is not a textbook; it contains no exercises, and is insufficiently detailed for that purpose. However, it could be used as a textbook, either in conjunction with the excellent texts of Weir (1996), Lange (1997) and Ott (1999), or by advanced students who will consult the cited literature for details.

The notes used at the Summer Course have been augmented by material from two lecture classes given at the University of Washington. A Special Topics class was given in January-March, 1999, and additional background on Markov chain Monte Carlo and Monte Carlo EM are included from that class. Some details were also first presented at a SEMSTAT workshop in Eindhoven in March 1999 (Thompson, 2000b). Although material has been added, the examples in Chapter 10 and on identity by descent under interference (section 11.2) were first presented at a Royal Statistical Society Meeting in London, in March 1999 (Thompson, 2000*a*). Versions of Figures 9.1, 10.1, 10.2, 10.6, and 10.7 first appeared in Thompson (2000*a*). However, the 11-chapter monograph follows closely the ten sessions of the Summer Course presentations, with chapter 2 being the only addition, providing statistical background with genetic examples. The order of Chapters 8 and 9 has been reversed from the Summer Course; a case can be made for either ordering.

A more basic Statistical Genetics class was given in Fall 1999, at University of Washington, and led to extensive revision of Chapters 1-4. It is hoped that the monograph can thus serve two purposes. For example, a more introductory course could cover of Chapters 1-4, with final material taken from sections 6.1, 6.2, 7.1, and 7.2. More advanced students could skip Chapters 1-2, skim Chapters 3-5, and study the later chapters more thoroughly.

I would like to thank Dr.Anant Godbole and Dr.JianPing Dong, for their excellent organization of the CBMS Regional Research Conference at Michigan Technical University. I am also grateful to the many students who attended this course, and to students attending the two University of Washington courses, for their helpful comments and criticisms. In particular, I would like to thank Eric Anderson, Nicky Chapman and Dr.Ellen Wijsman for help with LaTeX, BibTeX, Xfig, and GENEHUNTER, and for many discussions. I am grateful to Amy Anderson for her thorough and critical reading of Chapters 1 to 5, and to Eric Anderson, Dr.Erin Conlon, Dr.Mary Kuhner, Anne-Louise Leutenegger, and Jessica Maia, who all read and commented on other chapters.

Some of the MCMC work was undertaken in collaboration with Dr.Simon Heath. In particular, the implementation of the algorithm described in section 3.6 and the initial incorporation of the L-sampler of Heath (1997) into our M-sampler software to create the LM-sampler (section 10.6) are both due to Dr.Heath. Figures 1.1, 1.2, 3.4, 3.5, and 10.3, first appeared in Thompson and Heath (1999), and are also due to Dr.Heath. I am grateful to Dr.Heath for our continuing collaboration.

The CBMS Regional Research Conference was funded by NSF grant number 98-13767 to Dr.Jianping Dong and Dr.Anant Godbole of The Mathematical Sciences Department of Michigan Technical University, Houghton, MI.

Table of Notation

Since there are an insufficient number of user-friendly letters and symbols, some must be used for more than one purpose. However, for convenience, we summarize the principal usages here

e principal usag	ges nere	
Notation Usage		
Parameters		
heta	the general (set of) parameters of a model	
ρ	a recombination frequency parameter	
γ	a (trait) locus location	
Γ_M	a marker map; set of arker locations	
β	a trait model penetrance parameter	
r	number of multinomial outcomes (or phenotypes)	
p_1,\ldots,p_r	probabilities of multinomial outcomes	
k	number of alleles at a locus	
q_1,\ldots,q_k	population allele frequencies at this locus	
q	an allele frequency, often for a recessive allele	
$\hat{\psi}$	a kinship coefficient	
$\dot{\phi}$	chiasmata avoidance function	
$\kappa_i, i=0,1,2$	gene-identity probabilities	
Indices and		
\overline{i}	an index used primarily for individuals or meioses	
j	an index used primarily for alleles or loci	
k, k_i	a label for a gene	
L	a number of loci ordered on a chromosome	
m	a count, often of the number of meioses	
v	miscellaneous other counts, of genes for example	
n	sample size	
\overline{F}	father, or paternal, often as subscript	
M	mother, or maternal, often as subscript	
	also marker, as in marker data \mathbf{Y}_M	
Ν	Monte Carlo sample size	
	also (Chapter 5) the random number of chiasmata)	
au	an index of Monte Carlo or MCMC realizations	
au	a set of indices of latent variables	
\mathcal{D}	a set of indices of data observations	
Variables		
$A_1,, A_k$	the alleles at a locus	
\mathbf{Y} , value \mathbf{y}	the data random variables (usually phenotypes)	
\mathbf{Y}_M	phenotypes at marker loci, in linkage mapping	
\mathbf{Y}_T	trait phenotypes; $\mathbf{Y} = (\mathbf{Y}_T, \mathbf{Y}_M)$	
\mathbf{X} , value \mathbf{x}	latent variables	
\mathbf{X}^{\dagger}	a proposed value of \mathbf{X} in Monte Carlo sampling	
X*	a sampled or resampled value of X in Monte Carlo	
$\mathbf{G} = \{G_i\}$	the set of genotypes of individuals i	
	a genotype — a possible value of G_i	
<u>g</u>	a possible value of G1	

Notation	Usage
Variables continued	
$\mathbf{S} = \{S_{i,j}\}$	set of meiosis indicators for meioses i and loci j
$S_{\bullet,j}$	the vector of $S_{i,j}$ at given locus j
$S_{i,\bullet}$	the vector of $S_{i,j}$ at given meiosis i
$G_{\bullet,j}, G_{i,\bullet}$	similarly for genotypes, locus j , individual i
	similarly for phenotypes, locus j , individual i
$Y_{ullet,j}, Y_{i,ullet}$ $Y^{(j)}$	the data $\{Y_{\bullet,1}, Y_{\bullet,2}, \dots, Y_{\bullet,j}\}; \mathbf{Y} = Y^{(L)}$
$\mathbf{J} = \mathbf{J}(\mathbf{S})$	a gene <i>ibd</i> pattern, a function of \mathbf{S}
$I_1,, I_{L-1}$	the intervals between L ordered loci
$\mathbf{R} = (R_j; j = 1,, L - 1)$	the recombination indicators in intervals I_j
r = (103, 3 = 1,, 2 = 1)	a vector of recombination indices; value of \mathbf{R}
$C = (C_i; j = 1,, L - 1)$	the chiasmata presence/absence indicators in
$C = (C_j, j = 1,, L - 1)$	intervals I_{i}
a	
C T volue t	a vector of chiasma indices; value of C
T, value t	a count (often binomial)
t_j	a multinomial count, e.g. of latent genotypes;
	also (Chapter 5) a set of binary indicators
n_{jl}, n_{j}	multinomial data counts, of observable phenotypes
	or genotypes
m_j	multinomial counts, often of alleles
Functions and probabi	
Pr (The second s	probability, when not indexed by a parameter
$\Pr(E;\theta)$	probability of event E under model $ heta$
$P_{\theta}(\cdot)$	a probability distribution, indexed by $ heta$
$P^*(\cdot)$	a probability distribution, used for the sampling or
	resampling distribution in Monte Carlo methods
$\mathrm{E}_{m{ heta}}(\cdot)$	Expectation, under a model indexed by θ
$\Phi(\cdot)$	the standard Normal (Gaussian) cumulative
	distribution function
$I(\cdot)$	the indicator function of an event
$L(\theta) \text{ or } L_{\mathbf{y}}(\theta)$	the likelihood for parameter θ given data y
$L(\theta; \mathbf{Y})$	the likelihood function, considered also as a
	function of data random variables \mathbf{Y}
$\ell \text{ or } \ell(\theta)$	the log-likelihood function for parameter $ heta$
$K_n(\theta; \theta_0)$	Kullback-Leibler information in a sample size n
$K_{\mathbf{y}}(\theta;\theta_0)$	K-L information in latent \mathbf{X} given data \mathbf{y}
$H_{\mathbf{y}}(\theta;\theta_0)$	expected complete-data log-likelihood given
• • • •	$\mathbf{Y} = \mathbf{y}: \mathbf{E}_{\theta_0}(\log P_{\theta}(\mathbf{X}, \mathbf{Y}) \mid \mathbf{Y} = \mathbf{y})$
$R(\cdot)$ and $R^*(\cdot)$	cumulative probabilities of data used in computing
	probabilities on graphs or pedigrees
$Q(\cdot),Q^*(\cdot),Q^\dagger(\cdot)$	cumulative conditional probabilities of latent
~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~	variables given data on graphs or pedigrees
$h(\mathbf{X}^{\dagger};\mathbf{X})$	Hastings ratio for proposed $\mathbf{X}^{\dagger}$ when at state $\mathbf{X}$
,	
$egin{array}{l} q(\mathbf{X}^{\dagger};\mathbf{X}) \ a \end{array}$	proposal probability for $\mathbf{X}^{\dagger}$ when at state $\mathbf{X}$ the Metropolis-Hastings acceptance probability